

## BIO 365L Student Rig Cleanup Checklist

Date: \_\_\_\_\_

Rig #: \_\_\_\_\_

### Equipment:

1. \_\_\_ Microscope light off
2. \_\_\_ Video Monitor off
3. \_\_\_ Dagan Amplifier off
4. \_\_\_ Manometer off
5. \_\_\_ Log off your computer

\_\_\_\_\_ Initial to verify complete

### Gas diffuser(s):

1. \_\_\_ Turn off the gas at the wall valve
2. \_\_\_ Using a wash bottle, rinse off the air diffuser(s) first with deionized water, then with EtOH, then dry with a Kimwipe. Leave them hanging outside of the beakers to dry
3. \_\_\_ Briefly turn the gas back on (~ 10 sec) to blow out any remaining liquid then turn off.

\_\_\_\_\_ Initial to verify complete

### Perfusion system cleanup:

1. \_\_\_ Remove the slice from the chamber and discard. Put the harp back in the recording well
2. \_\_\_ Run 20 ml of EtOH through the perfusion system and the recording well (use the 50ml flask; you can increase the flow rate to hasten the process)
3. \_\_\_ Run 50 ml of deionized water through the perfusion system (use the 125 ml flask; you can increase the flow rate to hasten the process).
4. \_\_\_ When the perfusion lines are clean aspirate the remaining liquid from the recording well with the vacuum tube
5. \_\_\_ Turn off the vacuum to your rig by turning the valve labeled "VAC" so that it is perpendicular to the vacuum line.
6. \_\_\_ Remove the waste beaker underneath your rig and empty the waste into the sink. Replace the beaker.

\_\_\_\_\_ Initial to verify complete

### General:

1. \_\_\_ Empty your ACSF flasks, rinse with deionized water, hang them on drying rack
2. \_\_\_ All glass micropipettes disposed in Red Sharps container
3. \_\_\_ Using a wash bottle, rinse the 40x microscope objective with deionized water and blot dry with lens paper
4. \_\_\_ Rinse the wire on the pipette holder with EtOH then with deionized water

\_\_\_\_\_ Initial to verify complete